



Novel Genetic Variants Associated with Lumbar Spondylosis in Koreans : A Genome-Wide Association Study

Hyun Ah Kim, M.D., Ph.D.¹ Seong Gu Heo,² Ji Wan Park, Ph.D.,² Young Ok Jung, M.D., Ph.D.³

Division of Rheumatology,¹ Hallym University Sacred Heart Hospital, Anyang, Korea

Department of Medical Genetics,² Hallym University College of Medicine, Chuncheon, Korea

Division of Rheumatology,³ Gangnam Sacred Heart Hospital, Seoul, Korea

Objective : The aim of this study was to identify the susceptibility genes responsible for lumbar spondylosis (LS) in Korean patients.

Methods : Data from 1427 subjects were made available for radiographic grading and genome wide association studies (GWAS) analysis. Lateral lumbar spine radiographs were obtained and the various degrees of degenerative change were semi-quantitatively scored. A pilot GWAS was performed using the Affymetrix Genome-Wide Human single-nucleotide polymorphisms (SNPs), 500K array. A total of 352228 SNPs were analyzed and the association between the SNPs and case-control status was analyzed by stepwise logistic regression analyses.

Results : The top 100 SNPs with a cutoff p -value of less than 3.7×10^{-4} were selected for joint space narrowing, while a cutoff p -value of 6.0×10^{-4} was applied to osteophytes and the Kellgren-Lawrence (K-L) osteoarthritis grade. The SNPs with the strongest effect on disc space narrowing, osteophytes, and K-L grade were serine incorporator 1 (rs155467, odds ratio [OR]=17.58, $p=1.6 \times 10^{-4}$), stromal interaction molecule 2 (STIM1, rs210781, OR=5.53, $p=5 \times 10^{-4}$), and transient receptor potential cation channel, subfamily C (rs11224760, OR=3.99, $p=4.8 \times 10^{-4}$), respectively. Leucine-rich repeat-containing G protein-coupled receptor 4 was significantly associated with both disc space narrowing and osteophytes (rs1979400, OR=2.01, $p=1.1 \times 10^{-4}$ for disc space narrowing, OR=1.79, $p=3 \times 10^{-4}$ for osteophytes), while zinc finger and BTB domain containing 7C was significantly and negatively associated with both osteophytes and a K-L grade >2 (rs12457004, OR=0.25, $p=5.8 \times 10^{-4}$ and OR=0.27, $p=5.3 \times 10^{-4}$, respectively).

Conclusion : We identified SNPs that potentially contribute to the pathogenesis of LS. This is the first report of a GWAS in an Asian population.

Key Words : Spondylosis · Osteoarthritis · Single nucleotide polymorphism · Genome-wide association study.

• Received : October 5, 2016 • Revised : February 15, 2017 • Accepted : March 17, 2017

• Address for reprints : Young Ok Jung, M.D., Ph.D.

Division of rheumatology, Gangnam Sacred Heart Hospital, 1 Singil-ro, Yeongdeungpo-gu, Seoul 07441, Korea
Tel : +82-2-845-5305, Fax : +82-2-846-4669, E-mail : yjung@hallym.ac.kr

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INTRODUCTION

Low back pain (LBP) is an important public health problem and is associated with substantial societal costs in industrialized countries. Degenerative disease of the lumbar spine (lumbar spondylosis, LS), which is characterized radiologically by the presence of osteophytes, endplate sclerosis, and disc space narrowing¹³, is thought to be related to LBP. The prevalence of LS in Asian countries has scarcely been reported. However, ethnic differences in the prevalence of osteophytosis and disc degeneration have previously been noted²². Although it is widely known that the prevalence of LS increases with age, there exists a paucity of data on the risk factors for LS. LS shares many features with osteoarthritis (OA) of the peripheral joint, the most common form of arthritis affecting the elderly. Although the pathogenesis of OA has been associated with aging and environmental factors such as work activity, twin studies have revealed a significant genetic contribution, with a heritability estimate of 65% for hand OA²⁰. Recent reports have demonstrated the significant role that genetic contribution also plays in LS. A twin study showed that the heritability of LS was 74% for the lumbar spine and 73% for the cervical spine². Other studies also revealed significant heritability on the basis of magnetic resonance imaging (MRI) of the spine^{9,16}. The search for specific genes associated with OA and LS has since ensued. However, due to variations in the phenotype, as well as the severity of the disease and patient characteristics such as gender and age, the definitions of the disease have varied from study to study, leading to tremendous difficulty in identifying a causative genetic factor.

A large number of genetic association studies of OA have been conducted over the last decade, and a number of significant associations have been verified by a systematic review. For example, aggrecan 1 (*AGC1*) is associated with hand OA, asporin (*ASPN*) with hip and knee OA, and vitamin D receptor (*VDR*) with knee and spine OA¹⁵. However, the majority of the genes that were reported to have significant associations have never been replicated, making it difficult to discern the significance of these isolated significant associations in the pathogenesis of OA.

Complex conditions such as OA and LS are caused by numerous genetic and environmental factors, any one of which can have a relatively minor effect. Genome-wide association studies (GWAS) have provided a breakthrough in deciphering

the genetic influence in complex diseases, by examining hundreds of thousands of single-nucleotide polymorphisms (SNPs), thus enabling identification of SNPs implicated in a large number of robustly replicated loci of common traits.

Although GWAS has begun to unravel the genetic influence of peripheral joint OA²⁵, there has been little attempt as yet to do the same for LS. In addition, the results of an LS GWAS among an Asian population have not yet been reported. In this study, we sought to identify the susceptibility genes for LS in a GWAS among a community-based population of Korean adults.

MATERIALS AND METHODS

Study population

For this study, we selected a rural farming community (An-sung) in South Korea from among the populations incorporated into the ongoing prospective Korean Genome and Epidemiology Study (KoGES). The study methods have been described previously⁴. Briefly, the eligibility criteria included an age of 40–79 years, residence within the borders of the survey area for at least 6 months before testing, and the mental and physical ability to participate. Data relating to LBP were available for 4181 subjects for the years 2006 and 2007. Baseline demographic information was collected using a standard questionnaire during a face-to-face interview and included educational attainment, occupation, exercise, and co-morbidities.

Of the total number of eligible subjects, 2000 were randomly selected for spine radiography. No difference was found in the prevalence of LBP between those who underwent radiography and those who did not. After excluding 488 subjects who were unable to be evaluated due to poor film quality, and 85 patients whose genomic DNA was not obtained, data for 1427 subjects were available for radiograph grading and GWAS analysis. The study protocol was approved by the Ethics Committee of the KoGES, and written informed consent was obtained from each participant.

Radiographic evaluation of the lumbar spine

Lateral lumbar spine radiographs were taken according to a standard protocol with the film centered on the second lum-

bar vertebra. Lumbar radiographs were evaluated by a single observer. Each vertebral level from L1/2 to L4/5 was reviewed for the presence of radiographic features relating to degenerative change. Semi-quantitative scores were given for the following features using a reference atlas⁸⁾: presence and severity of anterior osteophytes (grade 0=none; grade 1=barely visible; grade 2=definite; grade 3=large), endplate sclerosis (grade 0=none; grade 1=present), and disc space narrowing (grade 0=none; grade 1=barely visible; grade 2=definite; grade 3=severe, bone to bone).

Additionally, the Kellgren-Lawrence (K-L) grading system was used for each vertebral level (grade 0=normal disc with no osteophytes; grade 1=slight anterior wear and osteophyte formation; grade 2=definite anterior wear and mild disc space narrowing with osteophyte formation; grade 3=moderate disc space narrowing with osteophytes and sclerosis; grade 4=large osteophytes, marked disc space narrowing, and sclerosis of vertebral end plates). LS was defined using one of the following criteria: 1) joint space narrowing \geq grade 2; 2) osteophytes \geq grade 2; 3) K-L grade \geq 2. Radiographs were read by a single reader who was an academically-based rheumatologist. Intra-observer reproducibility was assessed by re-evaluating 50 films within 1 week of the first reading. The reproducibility of intra-reader assessments was high (for osteophyte grading, $\kappa=0.89-0.93$; for endplate sclerosis, $\kappa=0.71-0.84$; for joint space narrowing, $\kappa=0.81-0.89$; and for K-L grading, $\kappa=0.69-0.80$, for various vertebral levels). Films allocated different grades at each of the two readings were adjudicated by consensus between the original reader and a second reader.

Genome-wide association study

Genomic DNA was isolated from peripheral blood mononuclear cells. We performed a pilot GWAS, typing cases and controls on a single platform using the Affymetrix Genome-Wide Human SNP 500K array chip (Affymetrix, Inc., Santa Clara, CA, USA). Genotype calls were determined by Bayesian robust linear modeling using the Mahalanobis distance algorithm¹⁴⁾. We sequentially discarded 38364 markers with a Hardy-Weinberg equilibrium *p*-value $<10^{-6}$, 17926 with genotype call rates below 95%, and 92050 with a minor allele frequency (MAF) of 0.01. This left 352228 SNPs available for subsequent analysis

Statistical analysis

The evaluation of the association between case-control status and each individual SNP was based on the odds ratio (OR) and *p*-values. Covariates used for multivariable-adjustment were age, sex, education, and body mass index. To optimize the joint effect of the SNPs, we conducted stepwise logistic regression analyses and also analyzed the main effects on the selected variables using a generalized linear model implemented with the R statistical software (ver. 2.14.2; R Core Development Team, Austria, Vienna).

RESULTS

Table 1 displays the clinical characteristics of the study population, with cases of spine OA defined as disc space narrowing, osteophytes, or K-L grade all \geq 2. Subjects with spine OA

Table 1. Demographic characteristics of the study subjects

	Disc space narrowing			Osteophyte			K-L grade		
	Case	Control	Total	Case	Control	Total	Case	Control	Total
Age (years)	64.7±0.3*	58.5±0.4	59.3±0.2	62.4±0.3*	56.3±0.3	59.5±0.2	61.2±0.3*	55.3±0.4	59.5±0.2
Female	132 (57.9)	699 (58.4)	831 (58.3)	369 (48.4)*	462 (69.6)	831 (58.2)	536 (52.1)*	293 (74.2)	829 (58.2)
BMI	24.1±0.2	24.4±0.1	24.3±0.1	24.3±0.1	24.3±0.1	24.3±0.1	24.3±0.1	24.2±0.2	24.3±0.1
Smoker	45 (19.7)	222 (18.6)	267 (18.7)	168 (22.1)*	100 (15.1)	268 (18.8)	215 (20.9)*	52 (13.2)	267 (18.8)
Alcohol	79 (34.7)*	505 (42.2)	584 (41.0)	323 (42.3)	261 (39.3)	584 (40.9)	429 (41.7)	155 (39.2)	584 (41.0)
Employed	216 (94.7)	1124 (93.8)	1340 (94.0)	723 (94.8)	617 (92.2)	1340 (93.9)	974 (94.7)	364 (92.2)	1338 (94.0)
Education \leq 6 years	64 (28.1)*	196 (16.4)	260 (18.2)	159 (20.8)*	101 (15.2)	260 (18.2)	207 (20.1)*	51 (12.9)	258 (18.1)

Values are presented as mean±standard deviation or number (%). Obesity=BMI \geq 27. *Denotes significant difference (*p*<0.05) compared to control. K-L: Kellgren-Lawrence, BMI: body mass index

were older, more likely to be male, and had a lower level of education compared to the control subjects.

Genome-wide association

We used logistic regression analysis to identify statistically significant associations with LS. For multiple comparisons, a Bonferroni-corrected p -value of 1.5×10^{-7} (0.05/320942) was

used. Since none of the SNPs reached the very conservative Bonferroni corrected value, the top 100 SNPs with a cutoff p -value less than 3.7×10^{-4} were selected for use in this GWAS for joint space narrowing while a cutoff p -value of 6.0×10^{-4} was applied for osteophytes and K-L grade. Tables 2–4 present the detailed characteristics of the SNPs, including gene name, rs identification number, position, MAF of the cases and controls, OR, and p -value. Each feature of LS was found to be as-

Table 2. Results of the genome-wide association study showing top 30 SNPs having the highest OR associated with ≥ 2 K-L grade

rs number	Chr	Gene	MAF	OR (95%CI)	Bonf p -value
rs11224760	11q22.1	NA	0.2299	3.99 (1.835–8.674)	0.00048
rs3753613	1p35.2	HCRTR1	0.2949	2.618 (1.577–4.347)	0.0002
rs10849640	12q24.23	NA	0.301	2.595 (1.534–4.39)	0.00038
rs11120305	1q41	PTPN14	0.3232	2.565 (1.552–4.24)	0.00024
rs2271933	1p35.2	HCRTR1	0.2824	2.476 (1.476–4.152)	0.00059
rs6569814	6q23.2	TAAR2 TAAR3	0.3197	2.385 (1.488–3.821)	0.0003
rs1164894	9q34.3	NA	0.3685	2.335 (1.522–3.582)	0.0001
rs10774756	12q24.21	LOC105369998	0.3997	2.212 (1.483–3.301)	0.0001
rs1473047	5q14.3	NA	0.3982	2.173 (1.46–3.234)	0.00013
rs10072084	5q14.3	NA	0.3966	2.135 (1.434–3.178)	0.00019
rs7966636	12q24.21	LOC105369998	0.3749	2.092 (1.374–3.183)	0.00057
rs197457	6q24.2	HIVEP2	0.3891	2.085 (1.403–3.1)	0.00028
rs3794214	12q24.31	ACADS	0.3825	2.039 (1.367–3.043)	0.00048
rs6868338	5q34	NA	0.392	1.984 (1.349–2.918)	0.0005
rs2834443	21q22.11	NA	0.4292	1.969 (1.368–2.835)	0.00027
rs9533738	13q14.11	LOC105370182	0.4776	1.89 (1.36–2.627)	0.00015
rs2378931	14q12	LOC105370438	0.4329	1.876 (1.326–2.654)	0.00038
rs10026693	4p16.1	SORCS2	0.4699	1.835 (1.325–2.542)	0.00026
rs2878620	4p16.1	SORCS2	0.4703	1.834 (1.324–2.54)	0.00027
rs3857194	4p16.1	SORCS2	0.471	1.794 (1.297–2.481)	0.00041
rs9884489	4p16.1	SORCS2	0.4706	1.794 (1.297–2.481)	0.00041
rs6833329	4p16.1	SORCS2	0.4706	1.794 (1.297–2.481)	0.00041
rs2937545	5p13.2	NA	0.4675	1.767 (1.28–2.441)	0.00055
rs873471	8p22	PSD3	0.488	0.6036 (0.4528–0.8047)	0.00058
rs2063076	8p22	PSD3	0.4941	0.6007 (0.451–0.8001)	0.00049
rs4667789	2q24.3	SCN2A SCN3A	0.4888	0.6002 (0.4491–0.8022)	0.00056
rs751217	8p22	PSD3	0.4941	0.599 (0.45–0.7974)	0.00045
rs7556825	2q24.3	SCN2A SCN3A	0.4968	0.5984 (0.4516–0.7929)	0.00035
rs2914908	5q23.1	NA	0.478	0.5889 (0.4398–0.7886)	0.00038
rs7576705	2q37.3	NA	0.486	0.5839 (0.4373–0.7796)	0.00026

SNPs : single-nucleotide polymorphisms, OR : odds ratio, K-L : kellygren-lawrence, CI : confidence interval, MAF : minor allele frequency, NA : not available

sociated with distinct SNPs. The SNPs that had the strongest effect on disc space narrowing included serine incorporator 1 (rs155467, OR=17.58, $p=1.6 \times 10^{-4}$), heat shock transcription factor 2 (rs563084, OR=14.48, $p=1.4 \times 10^{-4}$), cysteine-rich hydrophobic domain (rs1568512, OR=11.57, $p=4 \times 10^{-5}$), akirin 2 (rs2787938, OR=9.86, $p=8 \times 10^{-5}$), and fibroblast growth factor receptor 2 (rs11200052, OR=9.72, $p=9 \times 10^{-5}$), while those that had the most significant effect on osteophytes included stromal interaction molecule 2 (*STIM1*, rs210781, OR=5.53, $p=5 \times$

10^{-4}), protein kinase C and casein kinase substrate in neurons 2 (*PACSIN2*, rs738379, OR=5.37, $p=3.6 \times 10^{-4}$), and ubiquinol-cytochrome c reductase complex chaperone (rs6060373, OR=3.05, $p=8 \times 10^{-5}$). Transient receptor potential cation channel, subfamily C (rs11224760, OR=3.99, $p=4.8 \times 10^{-4}$), hypocretin (orexin) receptor 1 (rs3753613, OR=2.62, $p=2 \times 10^{-4}$), and coiled-coil domain containing 60 (rs10849640, OR=2.59, $p=3.8 \times 10^{-4}$) had the strongest effect on K-L grades >2. Leucine-rich repeat-containing G protein-coupled receptor 4 was

Table 3. Results of the genome-wide association study showing top 30 SNPs having the highest OR associated with ≥ 2 osteophyte

rs number	Chr	Gene	MAF	OR (95%CI)	Bonf p-value
rs210781	4p15.2	NA	0.1943	5.533 (2.107–14.53)	1.00E+00
rs738379	22q13.2	PACSIN2	0.1738	5.368 (2.133–13.51)	1.00E+00
rs2284097	22q13.2	PACSIN2	0.1737	5.358 (2.129–13.48)	1.00E+00
rs2038062	22q13.2	PACSIN2	0.1761	4.532 (1.901–10.8)	1.00E+00
rs6060373	20q11.22	UQCC1	0.2566	3.053 (1.756–5.306)	1.00E+00
rs6088791	20q11.22	UQCC1	0.2526	3.053 (1.754–5.312)	1.00E+00
rs1539581	1p13.2	NA	0.211	2.991 (1.653–5.411)	1.00E+00
rs6060369	20q11.22	UQCC1	0.2523	2.989 (1.714–5.211)	1.00E+00
rs2425062	20q11.22	UQCC1	0.2565	2.988 (1.716–5.204)	1.00E+00
rs8127664	21q22.3	NA	0.2411	2.852 (1.664–4.89)	1.00E+00
rs761166	22q13.31	PARVB	0.2511	2.707 (1.557–4.705)	1.00E+00
rs10998893	10q22.1	NA	0.2519	2.648 (1.52–4.612)	1.00E+00
rs17705721	4q34.3	NA	0.2458	2.638 (1.557–4.467)	1.00E+00
rs4911178	20q11.22	UQCC1	0.2573	2.549 (1.501–4.33)	1.00E+00
rs4911496	20q11.22	UQCC1	0.2575	2.544 (1.498–4.322)	1.00E+00
rs1570004	20q11.22	UQCC1	0.2568	2.542 (1.496–4.317)	1.00E+00
rs10503404	8p23.1	MSRA	0.2913	2.47 (1.546–3.946)	1.00E+00
rs12038162	1q44	SMYD3	0.2435	2.451 (1.513–3.97)	1.00E+00
rs761165	22q13.31	PARVB	0.2776	2.429 (1.491–3.957)	1.00E+00
rs930140	2q36.1	PAX3	0.3026	2.277 (1.466–3.535)	1.00E+00
rs11970088	6p22.3	NA	0.2904	2.262 (1.454–3.52)	1.00E+00
rs10958163	8q21.13	NA	0.308	2.198 (1.442–3.35)	1.00E+00
rs11578091	1q41	NA	0.3061	2.185 (1.426–3.347)	1.00E+00
rs10848193	12q24.33	NA	0.2966	2.128 (1.418–3.195)	1.00E+00
rs16934897	10p11.22	NA	0.3385	2.028 (1.373–2.995)	1.00E+00
rs10194645	2p25.3	NA	0.4203	1.932 (1.412–2.642)	1.00E+00
rs1380255	1q41	NA	0.3742	1.908 (1.342–2.711)	1.00E+00
rs2049164	2q34	ERBB4	0.3791	1.898 (1.359–2.651)	1.00E+00
rs12564579	1p36.21	KAZN	0.4042	1.85 (1.352–2.531)	1.00E+00
rs9360980	6q14.1	NA	0.4209	1.844 (1.346–2.526)	1.00E+00

SNPs : single-nucleotide polymorphisms, OR : odds ratio, MAF : minor allele frequency, CI : confidence interval, NA : not available

Table 4. Results of the genome-wide association study showing top 30 SNPs having the highest OR associated with ≥ 2 disc space narrowing

rs number	Chr	Gene	MAF	OR (95% CI)	Bonf <i>p</i> -value
rs155467	6q22.31	PKIB	0.08182	17.58 (3.979–77.71)	1.00E+00
rs155458	6q22.31	PKIB	0.08117	17.54 (3.97–77.47)	1.00E+00
rs563084	6q22.31	NA	0.08351	14.48 (3.653–57.41)	1.00E+00
rs1568512	4q12	CHIC2	0.08514	11.57 (3.61–37.08)	1.00E+00
rs2787938	6q15	NA	0.08875	9.858 (3.176–30.6)	1.00E+00
rs2754273	6q15	NA	0.08875	9.858 (3.176–30.6)	1.00E+00
rs6915593	6q15	NA	0.09005	9.826 (3.165–30.51)	1.00E+00
rs11200052	10q26.13	NA	0.1031	9.717 (3.126–30.21)	1.00E+00
rs3870374	8q24.13	NA	0.1226	7.942 (3.087–20.44)	1.00E+00
rs10459466	14q32.33	NA	0.1281	7.551 (2.54–22.45)	1.00E+00
rs11845269	14q32.33	NA	0.1296	6.731 (2.353–19.26)	1.00E+00
rs17113276	14q32.33	NA	0.1296	6.731 (2.353–19.26)	1.00E+00
rs10163015	15q26.2	MCTP2	0.1387	6.506 (2.595–16.31)	1.00E+00
rs4119133	4q32.1	NA	0.1618	6.003 (2.418–14.9)	1.00E+00
rs12499551	4q32.1	NA	0.1618	6.003 (2.418–14.9)	1.00E+00
rs4684126	3p25.2	IQSEC1	0.1069	5.891 (2.347–14.79)	1.00E+00
rs10133227	14q32.33	NA	0.1412	5.779 (2.273–14.69)	1.00E+00
rs8022729	14q32.33	NA	0.1428	5.769 (2.269–14.67)	1.00E+00
rs6536428	4q32.1	NA	0.143	5.763 (2.262–14.68)	1.00E+00
rs4446584	6q22.31	NA	0.1446	5.666 (2.328–13.79)	1.00E+00
rs6767561	3p25.2	IQSEC1	0.1071	5.615 (2.265–13.92)	1.00E+00
rs3742689	14q31.3	KCNK10	0.127	5.583 (2.305–13.52)	1.00E+00
rs279626	8q24.13	NA	0.1205	5.452 (2.236–13.29)	1.00E+00
rs4690943	4q32.1	NA	0.1433	5.413 (2.159–13.57)	1.00E+00
rs10108494	8q24.13	NA	0.1409	5.129 (2.22–11.85)	1.00E+00
rs10095460	8q24.13	NA	0.1412	5.123 (2.216–11.84)	1.00E+00
rs254411	5q14.1	NA	0.1445	5.066 (2.392–10.73)	1.00E+00
rs4540278	6q22.31	NA	0.1432	5.029 (2.126–11.9)	1.00E+00
rs279617	8q24.13	NA	0.1191	4.933 (2.056–11.84)	1.00E+00
rs7761112	6q22.31	NA	0.1432	4.85 (2.071–11.36)	1.00E+00

SNPs : single-nucleotide polymorphisms, OR : odds ratio, MAF : minor allele frequency, CI : confidence interval, NA : not available

significantly associated with both disc space narrowing and osteophytes (rs1979400, OR=2.01, $p=1.1 \times 10^{-4}$ for disc space narrowing, OR=1.79, $p=3 \times 10^{-4}$ for osteophytes), while zinc finger and BTB domain containing 7C was significantly and negatively associated with both osteophytes and a K-L grade > 2 (rs12457004, OR=0.25, $p=5.8 \times 10^{-4}$ for osteophytes, OR=0.27, $p=5.3 \times 10^{-4}$ for K-L grade). No SNP was found to be significantly associated with LS defined using all three criteria.

DISCUSSION

In this study, we investigated the association of clinical and genetic factors for LS defined by three features found on simple radiographs. The previously identified genetic risk factors for LS using a candidate approach included genes encoding extracellular matrix proteins expressed in the nucleus pulposus (inner structure) and annulus fibrosus (outer layer) of the disc, such as type IX collagen (*COL9A2* and *COL9A3*), *AGC1*,

and cartilage intermediate layer protein (*CILP*)¹⁵. These findings suggest that LS is caused by changes in the structural integrity of the intervertebral disc. Given that OA and LS are both degenerative diseases of the skeletal joints, and because articular cartilage and intervertebral discs share similar patterns of gene expression, it is expected that subjects with OA and LS have a similar genetic susceptibility. A systematic review of genetic associations for peripheral joint OA and degenerative disease of the spine revealed the difficulties related to complex phenotypes of these diseases. While many logical and reporting problems, including missing population details, multiple testing, and an over-reliance on subgroup analysis, were found, cases in which significant associations were replicated in independent studies were also identified¹⁵. Some of the genes thus identified are of functional importance. For example, *ASPN*, a member of the small leucine-rich proteoglycan family, inhibits in vitro chondrogenesis and the expression of *COL2A1* and *AGC1* through the inhibition of TGF- β signaling¹⁹.

A recent systematic search of the literature, including 52 studies, identified *ASPN* (D-repeat), *COL11A1* (rs1676486), growth differentiation factor 5 (*GDF5*) (rs143383), *SKT* (rs16924573), *THBS2* (rs9406328), and *MMP9* (rs17576) as genes associated with LS in humans defined by MRI with a moderate level of evidence⁵. However, the phenotype definition of lumbar disc degeneration was highly variable among the studies, including a decrease in disc signal intensity or disc height, disc bulges, disc herniations without specification of the symptoms, Modic changes, osteophytes, and lumbar spinal stenosis. In addition, the phenotype of disc degeneration varied between the initial and replication studies. As a result, the replications were inconsistent, and most of the associations were presented with a weak level of evidence.

We used simple x-ray for diagnose and grade lumbar spondylosis. The K-L grade is most popular grading system with classification into five grade scales (0–4) where K-L grade ≥ 2 is the conventional standard of the diagnosis⁸. Epidemiological studies showed that K-L grade was associated with the degree of low back pain in elderly subjects^{4,11}.

The variation in the identified genetic associations may reflect ethnic diversity as well as phenotypic heterogeneity. While *ASPN* was identified as a candidate gene for lumbar disc degeneration with a moderate level of epidemiological evidence among Asians, *GDF5* was instead identified among the

Northern European population²⁴. In addition, flaws inherent in a candidate gene approach, such as isolated analyses of disparate potential associations, may merely add to the growing repertoire of weak evidence. The genome-wide association study is based on the ‘common disease-common variation’ theory, which proposes that multiple common polymorphisms with a MAF of 0.5–1%, and with small effect sizes, might be predisposed to common disorders¹. Because more complex modes of inheritance involving multiple genes rather than a simple monogenic Mendelian pattern appear to be in effect for LS, the discovery of its associated genes is more likely with a GWAS than by an SNP analysis. A meta-analysis of 4 GWAS that addressed LS among 4683 individuals with European ancestries was reported by Williams et al.²³ LS was defined as disc space narrowing and the formation of osteophytes, as in our study. Among the four markers identified (rs17034687, rs2187689, rs7767277, and rs926849), the rs926849 marker located in the intronic region of Parkinson protein 2, the E3 ubiquitin protein ligase (*PARK2*) gene, on chromosome 6 remained strongest after adjusting for age and gender. The rs2187689 and rs7767277 markers were in strong linkage disequilibrium with proteasome subunit beta type 9 (large multifunctional peptidase 2) (*PSMB9*). Although we used the same 500K Affymetrix kit, the genes identified in our study did not overlap with those reported by Williams et al.²⁴. Again, the discrepancy may arise from ethnic differences between the study populations and the difference in phenotype definition.

The SNP of leucine-rich repeat-containing G protein-coupled receptor 4 (*LGR4*) was significantly associated with both disc space narrowing and osteophytes in our study. *LGR4* encodes a receptor for R-spondins (*RSPOs*), which play a pleiotropic role in normal development, and the development of cancer, as well as in the survival of adult stem cells through potentiation of Wnt signaling⁷. Knockout of *LGR4* or *RSPOs* in mice presents with severe developmental abnormalities, causing neonatal/embryonic lethality¹⁰. Recent studies showed that the *RSPO-LGR4* axis elevates the levels of Wnt receptors through direct inhibition of the ubiquitination of Wnt receptors or through interaction with intracellular signaling proteins to potentiate the Wnt pathways³. Increased Wnt/ β -catenin signaling is observed in the nucleus pulposus of dogs that suffer from premature intervertebral disc degeneration¹⁸, suggesting the role of Wnt/ β -catenin signaling, and possibly the *RSPO-LGR4* axis, in the pathogenesis of LS.

The SNP of the heat shock transcription factor 2 gene (*HSTF2*) was one of the SNPs most significantly associated with LS as defined by disc space narrowing. *HSTF2* specifically binds to the heat-shock promoter element and activates transcription for heat-shock response genes under conditions of heat or other stress. Previous studies showed that the formation of heterocomplexes between *HSTF1* and *HSTF2* leads to enhanced activity, which activates the hsp70 promoter, and that *HSTF2* was able to modulate the *HSTF1*-mediated expression of major heat shock protein genes¹². A previous study showed that *HSTF* expression was more frequent in clustered cells in both the annulus fibrosus and nucleus pulposus of herniated discs¹⁷. Because the K-L grade is accounted for more by osteophytes than by joint space narrowing, it is not surprising that genes associated with LS, as defined by the K-L grade, are different from those defined by disc space narrowing. The strongest association with K-L grade was observed with the SNP for transient receptor potential cation channel (*TRPC*), subfamily C, which encodes a receptor-activated calcium channel. In a study using OA chondrocytes, a correlation between the appearance of *TRPC6* and the state of de-differentiation of chondrocytes was identified⁶. Because the loss of a differentiated chondrocyte phenotype is one of the hallmarks of OA, as well as LS, the role of *TRPC6* in the process of disc degeneration is plausible. On the other hand, *TRPC6* was expressed in bone-derived cells and inhibitors of the TRP channel inhibited the effects of bradykinin-induced Ca^{2+} -influx, suggesting a role in bone metabolism²¹.

This is the first reported GWAS in an Asian population. In terms of phenotyping, we used disc space narrowing as well as osteophytosis and K-L grading to cover all aspects of degenerative change observable in LS. Limitations included a sample size too small to detect the true effect of SNPs. Although we did not identify the same loci reported in previous studies using a candidate gene approach, differences in ethnicity and phenotype definition may have affected the results. Further studies using larger Asian samples are warranted to establish if the gene variants identified in this study are associated with an increase in the risk of LS in Asian populations.

CONCLUSION

A GWAS was conducted to identify the susceptibility genes

responsible for lumbar spondylosis assessed by simple radiography. We identified SNPs that potentially contribute to the pathogenesis of LS. This is the first report of a GWAS in an Asian population.

PATIENT CONSENT

The patient provided written informed consent for the publication and the use of their images.

• Acknowledgements

This research was supported by a grant of the Korea Health Technology R&D Project through the Korea Health Industry Development Institute (KHIDI), funded by the Ministry of Health & Welfare, Republic of Korea (grant number : HI14C2248).

References

1. Altshuler D, Daly MJ, Lander ES : Genetic mapping in human disease. **Science** **322** : 881-888, 2008
2. Battié MC, Videman T, Gibbons LE, Fisher LD, Mannine H, Gill K : 1995 Volvo Award in clinical sciences. Determinants of lumbar disc degeneration. A study relating lifetime exposures and magnetic resonance imaging findings in identical twins. **Spine** **20** : 2601-2612, 1995
3. Carmon KS, Gong X, Yi J, Thomas A, Liu Q : RSP0-LGR4 functions via IQGAP1 to potentiate Wnt signaling. **Proc Natl Acad Sci U S A** **111** : E1221-E1229, 2014
4. Cho NH, Jung YO, Lim SH, Chung CK, Kim HA : The prevalence and risk factors of low back pain in rural community residents of Korea. **Spine (Phila Pa 1976)** **37** : 2001-2010, 2012
5. Eskola PJ, Lemmelä S, Kjaer P, Solovieva S, Männikkö M, Tommerup N, et al. : Genetic association studies in lumbar disc degeneration: a systematic review. **PLoS One** **7** : e49995, 2012
6. Gavenis K, Schumacher C, Schneider U, Einfeld J, Mollenhauer J, Schmidt-Rohlfing B : Expression of ion channels of the TRP family in articular chondrocytes from osteoarthritic patients: changes between native and in vitro propagated chondrocytes. **Mol Cell Biochem** **321** : 135-43, 2009
7. Glinka A, Dolde C, Kirsch N, Huang YL, Kazanskaya O, Inqelfinger D, et al. : LGR4 and LGR5 are R-spondin receptors mediating Wnt/ β -catenin and Wnt/PCP signalling. **EMBO Rep** **12** : 1055-1061, 2011
8. Lane NE, Nevitt MC, Genant HK, Hochberg MC : Reliability of new indices of radiographic osteoarthritis of the hand and hip and lumbar disc degeneration. **J Rheumatol** **20** : 1911-1918, 1993
9. Matsui H, Kanamori M, Ishihara H, Yudoh K, Naruse Y, Tsuji H : Familial

- predisposition for lumbar degenerative disc disease. A case-control study. **Spine (Phila Pa 1976)** **23** : 1029-1034, 1998
10. Mazerbourg S, Bouley DM, Sudo S, Klein CA, Zhang JV, Kawamura K, et al. : Leucine-rich repeat-containing, G protein-coupled receptor 4 null mice exhibit intrauterine growth retardation associated with embryonic and perinatal lethality. **Mol Endocrinol** **18** : 2241-2254, 2004
 11. Muraki S, Oka H, Akune T, Mabuchi A, En-yo Y, Yoshida M, et al. : Prevalence of radiographic lumbar spondylosis and its association with low back pain in elderly subjects of population-based cohorts: the ROAD study. **Ann Rheum Dis** **68** : 1401-1406, 2009
 12. Ostling P, Björk JK, Roos-Mattjus P, Roos-Mattjus P, Mezger V, Sistonen L : Heat shock factor 2 (HSF2) contributes to inducible expression of hsp genes through interplay with HSF1. **J Biol Chem** **282** : 7077-7086, 2007
 13. Pye SR, Reid DM, Smith R, Adams JE, Nelson K, Silman AJ, et al. : Radiographic features of lumbar disc degeneration and self-reported back pain. **J Rheumatol** **31** : 753-758, 2004
 14. Rabbee N, Speed TP : A genotype calling algorithm for affymetrix SNP arrays. **Bioinformatics** **22** : 7-12, 2006
 15. Ryder JJ, Garrison K, Song F, Hooper L, Skinner J, Loke Y, et al. : Genetic associations in peripheral joint osteoarthritis and spinal degenerative disease: a systematic review. **Ann Rheum Dis** **67** : 584-591, 2008
 16. Sambrook PN, MacGregor AJ, Spector TD : Genetic influences on cervical and lumbar disc degeneration: a magnetic resonance imaging study in twins. **Arthritis Rheum** **42** : 366-372, 1999
 17. Sharp CA, Roberts S, Evans H, Brown SJ : Disc cell clusters in pathological human intervertebral discs are associated with increased stress protein immunostaining. **Eur Spine J** **18** : 1587-1594, 2009
 18. Smolders LA, Meij BP, Onis D, Riemers FM, Bergknut N, Wubbolts R, et al. : Gene expression profiling of early intervertebral disc degeneration reveals a down-regulation of canonical Wnt signaling and caveolin-1 expression: implications for development of regenerative strategies. **Arthritis Res Ther** **29** : R23, 2013
 19. Song YQ, Cheung KM, Ho DW, Poon SC, Chiba K, Kawaguchi Y, et al. : Association of the asporin D14 allele with lumbar-disc degeneration in Asians. **Am J Hum Genet** **82** : 744-747, 2008
 20. Spector TD, Cicuttini F, Baker J, Loughlin J, Hart D : Genetic influences on osteoarthritis in women: a twin study. **BMJ** **312** : 940-943, 1996
 21. Suzuki Y, Kodama D, Goto S, Togari A : Involvement of TRP channels in the signal transduction of bradykinin in human osteoblasts. **Biochem Biophys Res Commun** **410** : 317-321, 2011
 22. van Tulder MW, Assendelft WJ, Koes BW, Bouter LM : Spinal radiographic findings and nonspecific low back pain. A systematic review of observational studies. **Spine (Phila Pa 1976)** **22** : 427-434, 1997
 23. Williams FM, Bansal AT, van Meurs JB, Bell JT, Meulenbelt I, Suri P, et al. : Novel genetic variants associated with lumbar disc degeneration in northern Europeans: a meta-analysis of 4600 subjects. **Ann Rheum Dis** **72** : 1141-1148, 2013
 24. Williams FM, Popham M, Hart DJ, de Schepper E, Biema-Zeinstra S, Hofman A, et al. : GDF5 single-nucleotide polymorphism rs143383 is associated with lumbar disc degeneration in Northern European women. **Arthritis Rheum** **63** : 708-712, 2011
 25. Zhai G, van Meurs JB, Livshits G, Meulenbelt L, Valdes AM, Soranzo N, et al. : A genome-wide association study suggests that a locus within the ataxin 2 binding protein 1 gene is associated with hand osteoarthritis: the Treat-OA consortium. **J Med Genet** **46** : 614-616, 2009